

SUGAR-SUBSTITUTED 2-AZETIDINONE CHOLESTEROL ABSORPTION INHIBITORS: ENHANCED POTENCY BY MODIFICATION OF THE SUGAR

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Abstract: A glucuronide conjugate of the potent 2-azetidinone cholesterol absorption inhibitor Sch 58235 was synthesized to confirm the structure of a metabolite isolated from in vivo sources. A series of 2-azetidinone glycosides was prepared via Schmidt trichloroimidate methodology. Enhanced cholesterol absorption inhibition was achieved by modification of the sugar moiety. © 1998 Elsevier Science Ltd. All rights reserved.

A combination of metabolism and SAR studies of the 2-azetidinone cholesterol absorption inhibitor 1, Sch 48461¹ led to the discovery of the highly potent analog 2, Sch 58235.² Analog 2 is 50 times as potent as 1 in reducing liver cholesterol esters when given orally in the seven day cholesterol fed hamster assay.

These studies also found that both 1 and 2 are rapidly converted in vivo to the corresponding glucuronides 3 and 4.3 Sufficient quantities of 3 and 4 were required to confirm the structures of metabolites isolated from in vivo sources.

Chemistry

We recently confirmed the structure of 3 by independent synthesis. The key glycosidal linkage of 3 was forged by a modified Mitsunobu coupling protocol by treatment of phenol 5 and the benzyl protected sugar 6 with tributylphosphine and 1,1'-(azodicarbonyl)dipiperdine (ADDP) in THF. Subsequent hydrogenolysis

with Pearlman's catalyst provided the β -glucuronide 3, identical by NMR, MS and HPLC to 3 isolated from in vivo sources.

We assumed that 4 could be readily prepared from 2 by similar Mitsunobu methodology. However, treatment of 2 with 6⁵, ADDP and tributylphosphine or 7⁶, ADDP and triphenylphosphine failed to afford 9. These disappointing results led us to explore other modes of glycosylation. We next explored the possibility of employing Schmidt trichloroimidate mediated glycosylation to prepare 4.⁷ Zinc bromide promoted Schmidt coupling of 2 and 8⁸ from 0 °C to reflux resulted in either no reaction or complex mixtures of products.⁹

We speculate that the failure to prepare 9 via Schmidt or Mitusunobu glycosylation of 2 may be due to interference of the benzylic OH group. The benzylic OH of 2 was protected as the corresponding acetate 10 by bisacetylation of 2 (Ac₂O, CH₂Cl₂, DMAP, Et₃N, 100%) and subsequent selective cleavage of the phenolic acetate (guanidine, MeOH, RT, 60%). Several attempts to promote glycoside formation of 10 and 8 with zinc bromide were unsuccessful. Coupling was finally achieved under more traditional Schmidt conditions, treatment of a -20 °C solution of 10 and 8 in CH₂Cl₂ with catalytic boron trifluoride etherate, which upon warming to room temperature afforded 11 in 95% yield. 11

Hydrolysis of the ester groups was accomplished by stirring a dilute solution of 11 in a mixture of Et₃N:H₂O:MeOH (1:1:3.5, 0.01 M with respect to 11) overnight to afford 4.¹² Alternately, the acetate groups of 11 could be selectively removed with KCN in methanol to provide the ester 12.¹³

Synthetic 4 was found to be identical by NMR, MS, and HPLC with 4 isolated from in vivo sources. 14 The stereochemistry at the anomeric center was determined to be beta by NMR studies, ($J_{anomeric\ H} = 7.3\ Hz$). The Schmidt trichloroimidate coupling protocol and described deprotection procedures were employed to prepare compounds 14 - 16 presented in Table 1.

Biological Results

The glucuronic acid 4 was found to be less potent than 2 when given orally in the seven day cholesterol fed hamster assay. As was observed with analogs of 3, variation of the C-6 substitutent of the sugar modulates cholesterol absorption inhibition. Both the ester 12 and the alcohol 14 are more potent than 4. Cholesterol absorption inhibition appears to be tolerant of modification of the sugar moiety as evidenced by the activity of the variety of functionalized sugars presented in the table. Of particular note is the disaccharide 16, which is 4 times as potent as 2. The disaccharide portion of 16 is found in naturally occurring and synthetic saponins such as 17, which are reported to be inhibitors of cholesterol absorption. Disaccharide 16 (ED₅₀: 0.01 mg/kg) appears to be considerately more potent than 17 (reported ED₅₀: 2 mg/kg). However, we have not assessed compounds 16 and 17 head to head in the cholesterol fed hamster assay.

Table 1: Cholesterol Absorption Inhibition Activity of Sugar Substituted 2-Azetidinones in Orally Dosed Seven Day Cholesterol Fed Hamsters.*

Liver Serum Cholesterol Cholesterol **Esters** Dose ED₅₀ Rl (% reduction) (mg/Kg/day) (% reduction) Compound (mg/Kg/day) H H -93 -43 0.04 Н ·· OH -50 4 -95 3 0.17 CO2H OAc 11 -53 -96 Ac 3 0.09 CO₂Me 12 Н 3 -48 -89 0.04 CO₂Me 13 -14 -58 NDa Αc 1.0 CH₂OAc ОН 14 Н -48 -92 1.0 0.08 -75 15 Ac -43 6.0 NDa CH₂OAc CH₂OAc 16 Н ··OH -36 -90 1.0 0.01 СН₂ОН CH₂OH

^{*}Compounds were evaluated in the cholesterol fed hamster model at the indicated dose (n = 6/group). All compounds were statistically different from the cholesterol fed control group (n = 6/group). The compounds were evaluated in separate studies hence, direct statistical comparisons among the compounds was not performed. (a) ND = Not Determined.

Conclusions

The synthetic β-glucuronide 4 was found to be identical with 4 isolated from in vivo sources. Although metabolism studies have shown that 2 is rapidly converted to 4 in vivo, 4 was found to be less potent than 2 when orally dosed in the seven day cholesterol fed hamster assay. Cholesterol absorption activity approaching that of 2 can be restored by variation of the C-6 substituent of the sugar. The ester 12 and alcohol 14 are more potent than the acid 4. The disaccharide 16, was the most potent cholesterol absorption inhibitor identified in this study ED₅₀: 0.01 mg/kg/day and is four times as potent as 2. Cholesterol absorption inhibition appears to be tolerant of substitution of the sugar moiety. However, since we are presently restricted to an in vivo assay, interpretation of the cholesterol absorption activity of the compounds in Table 1 may not be straight forward. The observed cholesterol absorption inhibition may be a reflection of a compound's bioavailability and/or ease of conversion to active metabolites and not its intrinsic cholesterol absorption activity.

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 1+MMR (400 MHz, CDCl₃): 7.26 (4H, m), 7.21 (2H, m), 7.01 (4H, m), 6.93 (2H, t, J = 8.4 Hz), 5.69 (1H, t, J = 6.7 Hz), 5.34 (2H, m), 5.29 (1H, m), 5.15 (1H, d, J = 7.2 Hz), 4.56 (1H, d, J = 2.1 Hz), 4.17 (1H, m),

- 3.73 (3H, s), 3.02 (1H, dt, J = 7.6, 2.3 Hz), 2.07 (14H, m), 1.85 (2H, m). HRMS (FAB): calcd for M + H: C₃₉H₄₀NO₁₃F₂, 768.2468, found 768.2460.
- 1-O-[4-[Trans-(3R,4S)-1-(4-fluorophenyl)-2-oxo-3-[3-[(S)-hydroxy-4-fluorophenyl)propyl]]-4-azetidinyl]-phenyl]-β-D-glucuronic Acid 4: 11 (5.08 g, 6.98 mmol) was dissolved in a mixture of methanol (127 mL) and triethylamine (127 mL) at room temperature. Water (445 mL) was added slowly via an addition funnel over 10 min in order to maintain a homogeneous solution. The resulting clear yellow solution was stirred over night. A small aliquot of the reaction mixture was quenched into a vial containing 1 M HCl and ethyl acetate. TLC (5% HOAc/20% MeOH/75% CH2Cl2) of the ethyl acetate layer indicated consumption of starting material. The methanol and triethylamine were removed on a rotory evaporator. The remaining solution was made acidic with 1M HCl, diluted with ethyl acetate, transfered to a separatory funnel and extracted with ethyl acetate. The extracts were combined, washed with 1M HCl, water and brine, dryed over anhydrous sodium sulfate and concentrated to a white solid 3.81 g (93%). The solid was redissolved in methylene chloride, concentrated onto enough silica such that a free flowing powder was obtained. The resulting powder was loaded onto a chromatography column packed with silica and 15% MeOH/CH2Cl2. Elution with 5% HOAc/15% MeOH/80% CH2Cl2 provided 2.95 g of pure 4 and 0.36 g of the slightly impure 4. The 2.95g of pure 4 was azeotroped first with toluene (3X) and then methanol (5X). The resultant solid was heated to 60°C overnight under vacuum to remove any residual solvent and provide the title compound as a white solid 2.6g (64%). ¹HMR (500 MHz, CD₃OD): 7.31 (4H, m), 7.26 (2H, m), 7.09 (2H, d, J = 8.5 Hz), 6.99 (4H, m), 4.96 (1H, anomeric, d, J = 7.3 Hz), 4.78 (1H, d, J = 2.1 Hz), 4.59 (1H, dd, J = 5.3, 6.5 Hz), 3.96 (1H, d, J = 9.7 Hz), 3.59 (1H, m), 3.48 (2H, m), 3.08 (1H, m), 1.88 (4H, m)CMR (400 MHz, CD₃OD): 172.56, 169.81, 163.72, 160.51, 159.14, 142.37, 135.36, 133.19, 128.92, 128.71, 126.08, 120.05, 118.61, 116.83, 102.27, 77.43, 76.66, 74.67, 73.86, 73.15, 62.13, 61.36, 37.60, 26.23. HRMS (FAB): calcd for M + H: C₃₀H₃₀NO₉F₂, 586.1889, found 586.1883. $[\alpha]^{21.4}$ °C_D -73° (5.88 mg/2 mL MeOH). HPLC: Metachem Inertsil C8 column (1.0 mL/min, solvent gradient 70% 0.2 M NH4Ac pH 6 Buffer/30% Acetonitrile gradient to 100% acetonitrile over 40 min.) Rt 4: 9 min. Rt 2: 40 min.
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